(Review)

Silica and mineral silicates causing autoimmune diseases

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ABSTRACT There are many environmental, occupational and medical substances that cause dysregulation of autoimmunity. Among these substances, the effects and causative mechanisms of silica particles and asbestos fibers are discussed in this review. Many epidemiological studies have shown a significant association between silica exposure and the occurrence of autoimmune diseases such as rheumatoid arthritis (RA), systemic sclerosis (SSc), systemic lupus erythematosus (SLE), and anti-neutrophil cytoplasm antibody (ANCA)-related vasculitis. Although the importance of NALP3 inflammasome as the initial immune reaction against silica particles has been recognized, the processes that form the various autoimmune diseases in silica-exposed patients remain unclear. Silica can activate various immune cells and cause an unbalance of regulatory T cells, responder T cells and T helper 17 cells, which might be key factors in understanding the silica-induced autoimmune alteration. In contrast, asbestos exposure shows a smaller association with autoimmune diseases. However, interesting findings have been reported regarding anti-endothelial and mesothelial cell antibodies detected in asbestos-exposed patients. Further investigations may contribute to elucidation of the mechanisms involved in environmental factor-induced modification of autoimmunity.

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Key words : Silica, Asbestos, Autoimmune diseases, Autoantibody

INTRODUCTION

It is well known that various environmental factors such as chemical agents, including silica, asbestos, metals, pesticides and solvents, physical agents such as ionizing radiation, ultraviolet radiation (sunlight), electric and magnetic fields, and biological agents that include infectious microorganisms, foods, molds, mycotoxins, and other toxins may influence

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the human immune system, particularly in regard to dysregulation of autoimmunity¹⁻⁷. Many epidemiological investigations and experimental studies have evaluated environmental factors that induce autoimmune diseases¹⁻⁷. In order to investigate the biological mechanisms by which these environmental factors cause autoimmune diseases, it is important to conduct risk and hazard

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analyses of environmental and occupational substances, as well as safeguard the health of workers in various occupational situations and people living in hazardous residential areas¹⁻⁷. In addition to these aims, these investigations may be useful in determining the causative mechanisms of various autoimmune diseases, among which the pathophysiological and immunological aspects are well known, but the causative mechanisms remain unclear⁸⁻¹⁰.

In this review, we investigate silica and mineral silicate, a form of asbestos, by evaluating and discussing epidemiological and experimental findings in an effort to better understand environmental factor-induced autoimmune disorders.

SILICA

Epidemiological studies

There are many reports describing the epidemiological significance of silica exposure and complicated autoimmune diseases. Caplan syndrome, complicated pneumoconiosis, and rheumatoid arthritis (RA) were initially reported in 1953 by Caplan¹¹⁾. Although it was thought that the causes of pneumoconiosis were not confined to silica, and included coal and asbestos, subsequent reports suggested that silica is the most common cause of this syndrome¹²⁻¹⁷⁾. Another important syndrome involves complications of silicosis and systemic sclerosis, and is known as Erasmus syndrome. In 1957 Erasmus described an apparently high prevalence of progressive systemic sclerosis (PSS) / systemic sclerosis (SSc) in Witwatersrand gold miners exposed to dust containing free silica¹⁸⁾.

Recent epidemiological investigations involving meta-analysis of factors associated with exposure to silica indicate that the relative risk for RA in the silica-exposed population is more than three times higher than that of non-exposed people^{19–22)}. Meta-analysis has also been used to investigate the

association between silica and SSc²²⁻²³⁾. All of these studies indicate the significant high risk of SSc in the silica-exposed population²²⁻²³⁾. Studies have also been performed regarding the association between silica exposure and systemic lupus erythematosus (SLE)^{22, 24-28)}. A positive association was reported in each of these studies and a high risk was indicated. In addition, case reports and case-control studies have been conducted concerning complications of anti-neutrophil cytoplasmic antibody (ANCA)related vasculitis²⁹⁻⁴⁰⁾. These investigations revealed a higher prevalence of ANCA in silicosis patients²⁹⁻⁴⁰⁾.

The overall findings of these epidemiological studies indicate with certainty that silica exposure alters human autoimmune tolerance and causes various autoimmune diseases.

Mechanistic investigations

Investigations concerning the mechanisms by which silica exposure affects the regulation of autoimmunity suggest that inflammasomes are very important⁴¹⁻⁴⁵. They contribute toward the innate immunity against various external factors (including silica, asbestos, bacteria and their toxins) and internal factors (including crystalline derived from uric acid and cholesterol) that represent danger signals which need to be handled as the first step in a series of successive phenomena. The inflammasomes are large multi-molecular complexes composed of caspase-1, PYCARD/ASC (apoptosisassociated speck-like protein containing caspase recruitment domain), and NALPs (a type of NODlike receptors) that induce the secretion of mature interleukin (IL)-1 β and IL-18 (pro-inflammatory cytokines) from antigen-presenting cells (APC)⁴⁶ ⁻⁵⁰⁾. Although the activation of inflammasomes can contribute to the formation of lung fibrosis in silicosis and asbestosis, two typical forms of pneumoconiosis 51-53, the long-term and gradual development of dysregulation of autoimmunity may require additional cellular and molecular mechanisms involving various cell-to-cell, cytokineto-cytokine, and molecule-to-molecule interactions.

There are many players in the development of autoimmune diseases when considering cell-to-cell interactions, such as APCs to present self-antigens. T cells to recognize them, B cells to produce antibodies, regulatory T cells (Treg) to control / inhibit the self-reactivity, and T helper (Th) 17 cells to promote hyper-reactivity to self-antigens $^{54-58)}$. The cytokine network plays a key role during any alteration or disturbance of the control of immune tolerance⁵⁹⁻⁶³⁾. As mentioned above, IL-1 β and IL-18 are produced from APC⁴⁶⁻⁵⁰⁾. IL-1 α is also secreted from monocyte-macrophage-APC cells and has various biological effects on immune, neurological and hematological conditions. IL-1 α can expand Th1 cells to produce interferon (IFN)- γ . IFN- γ stimulates genes such as interferon regulatory factor 1 (IRF1) as the transcription factor to activate immune reactions^{64, 65)}. The networks then enlist nuclear factor- κB (NF κB) to activate self-antigen reacting immune cells such as B cells and T cells.

Unfortunately, it is not fully understood how the silica particles activate these networks through cell, cytokine and molecular interactions. The activation of inflammasomes is only the initial step of a series of cascading events that induce dysregulation of autoimmunity^{46–50}.

Silica-induced chronic activation of T cells

In addition to the adjuvant effects of silica that present various self-antigens to APCs with sufficient size as antigens, silica particles can also activate responder Th1 cells and Treg as we reported previously⁶⁶⁻⁷⁰⁾. Evidence of silica-induced activation was found with the expression of CD69, an early activation T-cell marker, when T cells were cultured with silica particles *in vitro* and resulted in a remarkably higher expression of Programed Death-1 (PD-1) genes in peripheral blood T cells compared to those derived from healthy donors (HD), as well as higher soluble IL-2 receptor (R) in the serum of silicosis patients compared to HD^{66-70} .

Silica also changes activation-induced cell death in stimulated helper T cells by altering transcription of the Fas/CD95 cell death receptor gene, and results in the longer survival of these T cells^{71–75)}. Soluble Fas (sFas), an alternative spliced variant of the Fas gene, was more highly expressed relative to the wild-type Fas transcript in T cells from silicosis patients compared to those from HD^{72–74)}. In addition, sFas was elevated in the serum of silicosis patients⁷⁵⁾. sFas prevents activation-induced cell death in stimulated T cells, resulting in the long-term survival of these T cells in which selfrecognizing clones may be present^{72, 76–79)}.

In contrast to results obtained for responder T cells, Treg expressed Fas/CD95 to a much higher level when it was chronically activated, and Fas/CD95 expression was higher in Treg derived from the peripheral blood of silicosis patients compared to HD⁷⁰. Furthermore, peripheral blood mononuclear cells from HD were cultured with silica particles and results indicated that forkhead box P3 (FoxP3) gene positive Treg was decreased⁷⁰. The overall findings indicate an unbalance, increase of responder T cells, and decrease of Treg that may form the base condition in silicosis patients for the development of dysregulation of autoimmunity^{76–79}.

Th17 and silicosis

A consideration of autoimmune diseases indicates that another T-cell type, Th17, plays an important role. The balance between Treg and Th17 is critical for the development of autoimmune diseases; Treg can be induced by transforming growth factor (TGF) β and IL-2, and if IL-6 is also present, T cells are polarized to Th17 rather than Treg. The decrease of Treg and increase of Th17 may be the important factors that induce autoimmune diseases^{80–85)}.

However, the relationship between silica exposure and alteration of Th17 cells remains unclear. It has been reported that Th17 can regulate silica-induced lung inflammation through an IL-1 β -dependent mechanism⁸⁰⁻⁸⁵. As mentioned above, pulmonary inflammation and development of fibrosis in the lung may initially be induced by activation of the NALP3 inflammasome, and the previous findings therefore seem reasonable. This would ensure direct and recurrent contact between Th17 cells and inhaled remaining silica particles in the lung, as well as in lymph nodes, and may alter the features of Th17 cells. Additionally, chronic inflammation and progression of lung fibrosis may alter cytokine conditions toward an IL-6-dominant state in silicosis patients to result in a Th17-dominant and Tregreduced situation in these patients⁸⁰⁻⁸⁵⁾. Factoranalysis of our preliminary data indicated that the serum IL-6 level is included in the same factor with the titer of anti-nuclear antibody (ANA) and serum level of ANCA when the levels of various cytokines and titers of auto-antigens were analyzed in silicosis patients. This may suggest that the level of IL-6 (as well as TGF- β) is a key factor for the detection of silicosis patients prone to dysregulation of autoimmunity⁸⁰⁻⁸⁵⁾.

MINERAL SILICATE, ASBESTOS

We would like to turn our attention now to the mineral silicate, a form of asbestos. The core molecule of asbestos comprises Si and O, and is therefore similar to silica. The physical features of asbestos differ completely from those of silica, and partly result from the fact that silica comprises particulate matter, whereas asbestos is fibrous and includes various other minerals such as magnesium and iron. Silica and asbestos exposure causes pneumoconiosis, i.e., lung fibrosis, and both are categorized as a Group 1 carcinogen by the International Agency for Cancer Research (IARC)^{86, 87)}. However, evidence that

asbestos causes dysregulation of autoimmunity is insufficient, and contrasts with the affirmative evidence found for silica^{88–95)}. Several case-control studies have shown a positive concern for the role of asbestos exposure in the development of RA^{88–95)}. Furthermore, some studies have shown immune enhancement or activation of autoimmunity defined by elevated immunoglobulins, rheumatoid factors, and ANA or ANCA following asbestos exposure without any confirmed symptoms of autoimmune diseases^{88–95)}.

Some interesting investigations have been published regarding the impact of asbestos $exposure^{96-100)}$. These reports indicated that asbestos exposure induces production of autoantibodies against specific cell types, and complement previous reports showing the presence of anti-endothelial cell antibodies in vasculitis, SSc and SLE¹⁰¹⁻¹⁰³⁾. Similar to findings concerning these antibodies, reports indicate that anti-fibroblast antibodies (AFAs) and anti-mesothelial cell antibodies (MCAAs) are present respectively in the serum of mice and humans that have been exposed to "Libby Amphibole (LA)"⁹⁶⁻¹⁰⁰⁾. LA is a mixture of amphibole fibers that contaminated the vermiculite that was mined outside of Libby, Montana, U.S.A. The interesting aspects of these antibodies include collagen production caused by binding with target cells such as AFAs to fibroblasts, and MCAAs to mesothelial $cells^{96-100)}$. The overall results suggest that asbestos exposure and production of these antibodies may play causative roles in asbestos-induced fibrosis formation in the lungs and pleura⁹⁶⁻¹⁰⁰.

Although these findings were not consistent with systemic autoimmune diseases, it is very interesting that antibodies against these specific cell types may contribute to asbestos-induced disease formation such as fibrosis⁹⁶⁻¹⁰⁰⁾.

CONCLUSION

There are many reviews of environmental

substances, drugs and chemicals that cause dysregulation of autoimmunity¹⁻⁷⁾. Various drugs including hydralazine, quinidine and procainamide are associated with the development of $SLE^{1-7)}$. Silica and silicone (breast augmentation) are known to be related to $SSc^{1-7)}$. In addition, certain organic chemicals such as aromatic hydrocarbons, including toluene, benzene and xylene, aliphatic chlorinated hydrocarbons such as vinyl chloride, trichloroethylene and perchloroethylene, epoxy resins, and metaphenylenediamine are known to cause SSc following environmental or occupational $exposure^{1-7)}$.

In this review, we only focused on silica and silicate, a form of asbestos, since silica is known as the most frequent environmental factor causing dysregulation of autoimmunity¹⁻⁷⁾. The summarized findings detailed in this review are presented in Fig. 1. New cases of pneumoconiosis, including silicosis and asbestosis, have been rapidly decreasing in the previous two to three decades because of improvement in occupational environments



Fig. 1. Schematic presentation of the biological effects of silica and mineral silicate, a form of asbestos, detailing fibrous changes in the lung, autoimmune diseases in silica-exposed patients, cancer in asbestos-exposed patients, immune stimulation and activation to yield various auto-antibodies, complications of autoimmune diseases in silica-exposed patients, and production of antibodies against endothelial and mesothelial cells in asbestos-exposed patients through activation of inflammasomes.

and material handling procedures. However, we now have to consider low-level exposure to environmental factors causing autoimmune diseases, as well as individual factors such as HLA types and single nucleotide polymorphism (SNP) in certain genes, including the epigenetic status of various genes, since these situations may influence the occurrence of systemic autoimmune diseases due to the relationship between exposure to environmental factors and individual situations.

Further studies are required to elucidate the detailed pathogenic mechanisms involved in environmental factor-induced dysregulation of autoimmunity. These investigations may contribute to the development of tools for the prevention and treatment of various autoimmune disorders.

CONFLICT OF INTEREST

All authors have nothing to declare conflict of interest regarding this study.

REFERENCES

- Miller FW, Alfredsson L, Costenbader KH, Kamen DL, Nelson LM, Norris JM, De Roos AJ: Epidemiology of environmental exposures and human autoimmune diseases: findings from a National Institute of Environmental Health Sciences Expert Panel Workshop. J Autoimmun 39: 259-271, 2012. doi: 10.1016/ j.jaut.2012.05.002.
- 2) D'Cruz D: Autoimmune diseases associated with drugs, chemicals and environmental factors. Toxicol Lett 112-113: 421-32, 2000
- 3) Cooper GS, Miller FW, Germolec DR: Occupational exposures and autoimmune diseases. Int Immunopharmacol 2: 303-313, 2002
- 4) Hess EV: Environmental chemicals and autoimmune disease: cause and effect. Toxicology 181-182: 65-70, 2002
- 5) Van Loveren H, Vos JG, Germolec D, Simeonova PP, Eijkemanns G, McMichael AJ: Epidemiologic associations between occupational and environmental exposures and autoimmune disease: report of a meeting to explore current evidence and identify research needs.

Int J Hyg Environ Health 203: 483-495, 2001

- 6) Powell JJ, Van de Water J, Gershwin ME: Evidence for the role of environmental agents in the initiation or progression of autoimmune conditions. Environ Health Perspect 10785: 667-672, 1999
- 7) Mayes MD: Epidemiologic studies of environmental agents and systemic autoimmune diseases. Environ Health Perspect 107S5: 743-748, 1999
- Khamashta MA, Manuel Ramos-Casals M (eds). Autoimmune Diseases: Acute and Complex Situations. Berlin, Germany. Springer. 2011
- 9) Mackay IR, Rose NR (eds). The Autoimmune Diseases (Kindle). Cambridge, Massachusetts, U.S.A. Academic Press. 2013
- Brenner KJ (ed). Autoimmune Diseases: Symptoms, Diagnosis and Treatment. New York, U.S.A. Nova Science Pub Inc. 2011
- Caplan A: Certain unusual radiological appearances in the chest of coal-miners suffering from rheumatoid arthritis. Thorax 8: 29-37, 1953. doi:10.1136/thx.8.1.29.
- Constantinidis K: Pneumoconiosis and Rheumatoid arthritis (Caplan's syndrome). Br J Clin Pract 31: 25-31, 1977
- Uber CL, McReynolds RA: Immunotoxicology of silica. Crit Rev Toxicol 10: 303-319, 1982
- Green FH, Laqueur WA: Coal workers' pneumoconiosis. Pathol Annu 15: 333-410, 1980
- Ondrasík M. Caplan's syndrome: Baillieres Clin Rheumatol 3: 205-210, 1989
- Kelly CA: Rheumatoid arthritis: classical rheumatoid lung disease. Baillieres Clin Rheumatol 7: 1-16, 1993
- 17) Schreiber J, Koschel D, Kekow J, Waldburg N, Goette A, Merget R: Rheumatoid pneumoconiosis (Caplan's syndrome). Eur J Intern Med 21: 168-72, 2010. doi: 10.1016/j.ejim.2010.02.004.
- 18) Erasmus LD: Scleroderma in gold miners on the Witwatersrand with particular reference to pulmonary manifestations. S Afr J Lab Clin Med. 3: 209-231, 1957
- Khuder SA, Peshimam AZ, Agraharam S: Environmental risk factors for rheumatoid arthritis. Rev Environ Health 17: 307-315, 2002
- 20) Stolt P, Källberg H, Lundberg I, Sjögren B, Klareskog L, Alfredsson L; EIRA study group: Silica exposure is associated with increased risk of developing rheumatoid arthritis: results from the Swedish EIRA study. Ann Rheum Dis 64: 582-586, 2005

- 21) Stolt P, Yahya A, Bengtsson C, Källberg H, Rönnelid J, Lundberg I, Klareskog L, Alfredsson L; EIRA Study Group: Silica exposure among male current smokers is associated with a high risk of developing ACPA-positive rheumatoid arthritis. Ann Rheum Dis 69: 1072-1076, 2010. doi: 10.1136/ard.2009.114694.
- 22) Makol A, Reilly MJ, Rosenman KD: Prevalence of connective tissue disease in silicosis (1985-2006) - a report from the state of Michigan surveillance system for silicosis. Am J Ind Med 54: 255-262, 2011. doi: 10.1002/ ajim.20917.
- 23) McCormic ZD, Khuder SS, Aryal BK, Ames AL, Khuder SA: Occupational silica exposure as a risk factor for scleroderma: a meta-analysis. Int Arch Occup Environ Health 83: 763-769, 2010. doi: 10.1007/s00420-009-0505-7.
- 24) Parks CG, Cooper GS, Nylander-French LA, et al.: Occupational exposure to crystalline silica and risk of systemic lupus erythematosus: a population-based, casecontrol study in the southeastern United States. Arthritis Rheum 46: 1840-1850, 2002
- 25) Finckh A, Cooper GS, Chibnik LB, Costenbader KH, Watts J, Pankey H, Fraser PA, Karlson EW: Occupational silica and solvent exposures and risk of systemic lupus erythematosus in urban women. Arthritis Rheum 54: 3648-3654, 2006
- 26) Cooper GS, Wither J, Bernatsky S, Claudio JO, Clarke A, Rioux JD. CaNIOS GenES Investigators, Fortin PR: Occupational and environmental exposures and risk of systemic lupus erythematosus: silica, sunlight, solvents. Rheumatology (Oxford) 49: 2172-2180, 2010. doi: 10.1093/rheumatology/keq214.
- 27) Conrad K, Mehlhorn J, Lüthke K, Dörner T, Frank KH: Systemic lupus erythematosus after heavy exposure to quartz dust in uranium mines: clinical and serological characteristics. Lupus 5: 62-69, 1996
- 28) Brown LM, Gridley G, Olsen JH, Mellemkjaer L, Linet MS, Fraumeni JF Jr: Cancer risk and mortality patterns among silicotic men in Sweden and Denmark. J Occup Environ Med 39: 633-638, 1997
- 29) Gregorini G, Ferioli A, Donato F, Tira P, Morassi L, Tardanico R, Lancini L, Maiorca R: Association between silica exposure and necrotizing crescentic glomerulonephritis with p-ANCA and anti-MPO antibodies: a hospital-based case-control study. Adv Exp Med Biol 336: 435-440, 1993

- Tervaert JW, Stegeman CA, Kallenberg CG: Silicon exposure and vasculitis. Curr Opin Rheumatol 10: 12-17, 1998
- 31) Gregorini G, Tira P, Frizza J, D'Haese PC, Elseviers MM, Nuyts G, Maiorca R, De Broe ME: ANCAassociated diseases and silica exposure. Clin Rev Allergy Immunol 15: 21-40, 1997
- 32) Wichmann I, Sanchez-Roman J, Morales J, Castillo MJ, Ocaña C, Nuñez-Roldan A: Antimyeloperoxidase antibodies in individuals with occupational exposure to silica. Ann Rheum Dis 55: 205-207, 1996
- 33) Hogan SL, Satterly KK, Dooley MA, Nachman PH, Jennette JC, Falk RJ. Glomerular Disease Collaborative Network: Silica exposure in anti-neutrophil cytoplasmic autoantibody-associated glomerulonephritis and lupus nephritis. J Am Soc Nephrol 12: 134-142, 2001
- 34) Brener Z, Cohen L, Goldberg SJ, Kaufman AM: ANCAassociated vasculitis in Greek siblings with chronic exposure to silica. Am J Kidney Dis 38: E28, 2001
- 35) Saeki T, Fujita N, Kourakata H, Yamazaki H, Miyamura S: Two cases of hypertrophic pachymeningitis associated with myeloperoxidase antineutrophil cytoplasmic autoantibody (MPO-ANCA)-positive pulmonary silicosis in tunnel workers. Clin Rheumatol 23: 76-80, 2004
- 36) Mulloy KB: Silica exposure and systemic vasculitis. Environ Health Perspect 2003 Dec;111(16):1933-1938, 2003
- 37) Hogan SL, Cooper GS, Savitz DA, Nylander-French LA, Parks CG, Chin H, Jennette CE, Lionaki S, Jennette JC, Falk RJ: Association of silica exposure with antineutrophil cytoplasmic autoantibody small-vessel vasculitis: a population-based, case-control study. Clin J Am Soc Nephrol 2: 290-299, 2007
- 38) Bartůnková J, Pelclová D, Fenclová Z, Sedivá A, Lebedová J, Tesar V, Hladíková M, Klusácková P: Exposure to silica and risk of ANCA-associated vasculitis. Am J Ind Med 49: 569-576, 2006
- 39) Shibuya H, Sano H, Osamura K, Kujime K, Hara K, Hisada T: Microscopic polyangiitis accompanied by pleuritis as the only pulmonary manifestation of occupational silica exposure. Intern Med 49: 925-929, 2010
- 40) Chen M, Kallenberg CG: The environment, geoepidemiology and ANCA-associated vasculitides. Autoimmun Rev 9: A293-298, 2010. doi: 10.1016/ j.autrev.2009.10.008.

- 41) Hornung V, Bauernfeind F, Halle A, Samstad EO, Kono H, Rock KL, Fitzgerald KA, Latz E: Silica crystals and aluminum salts activate the NALP3 inflammasome through phagosomal destabilization. Nat Immunol 9: 847-856, 2008. doi: 10.1038/ni.1631.
- 42) Kuroda E, Ishii KJ, Uematsu S, Ohata K, Coban C, Akira S, Aritake K, Urade Y, Morimoto Y: Silica crystals and aluminum salts regulate the production of prostaglandin in macrophages via NALP3 inflammasome-independent mechanisms. Immunity 34: 514-526, 2011. doi: 10.1016/j.immuni.2011.03.019.
- 43) Shannahan JH, Ghio AJ, Schladweiler MC, Richards JH, Andrews D, Gavett SH, Kodavanti UP: Transcriptional activation of inflammasome components by Libby amphibole and the role of iron. Inhal Toxicol 24: 60-69, 2012. doi: 10.3109/08958378.2011.633942.
- 44) Sandberg WJ, Låg M, Holme JA, Friede B, Gualtieri M, Kruszewski M, Schwarze PE, Skuland T, Refsnes M: Comparison of non-crystalline silica nanoparticles in IL-1β release from macrophages. Part Fibre Toxicol 9: 32, 2012. doi: 10.1186/1743-8977-9-32.
- 45) Burton L, Paget D, Binder NB, Bohnert K, Nestor BJ, Sculco TP, Santambrogio L, Ross FP, Goldring SR, Purdue PE: Orthopedic wear debris mediated inflammatory osteolysis is mediated in part by NALP3 inflammasome activation. J Orthop Res 31: 73-80, 2013. doi: 10.1002/jor.22190.
- 46) Pétrilli V, Dostert C, Muruve DA, Tschopp J: The inflammasome: a danger sensing complex triggering innate immunity. Curr Opin Immunol 19: 615-622, 2007
- 47) Franchi L, Park JH, Shaw MH, Marina-Garcia N, Chen G, Kim YG, Núñez G: Intracellular NOD-like receptors in innate immunity, infection and disease. Cell Microbiol 10: 1-8, 2008
- Lee MS, Kim YJ: Pattern-recognition receptor signaling initiated from extracellular, membrane, and cytoplasmic space. Mol Cells 23: 1-10, 2007
- 49) Lamkanfi M, Kanneganti TD, Franchi L, Núñez
 G: Caspase-1 inflammasomes in infection and inflammation. J Leukoc Biol 82: 220-225, 2007
- 50) Lee MS, Kim YJ: Signaling pathways downstream of pattern-recognition receptors and their cross talk. Annu Rev Biochem 76: 447-480, 2007
- 51) Heppleston AG: Silica and asbestos: contrasts in tissue response. Ann NY Acad Sci 330: 725-744, 1979
- 52) deShazo RD: Current concepts about the pathogenesis of

silicosis and asbestosis. J Allergy Clin Immunol 70: 41-49, 1982

- 53) Doll NJ, Stankus RP, Barkman HW: Immunopathogenesis of asbestosis, silicosis, and coal workers' pneumoconiosis. Clin Chest Med 4: 3-14, 1983
- 54) De Carli M, D'Elios MM, Zancuoghi G, Romagnani S, Del Prete G: Human Th1 and Th2 cells: functional properties, regulation of development and role in autoimmunity. Autoimmunity 18: 301-308, 1994
- 55) Shevach EM, McHugh RS, Piccirillo CA, Thornton AM: Control of T-cell activation by CD4+ CD25+ suppressor T cells. Immunol Rev 182: 58-67, 2001
- 56) Chen W, Wahl SM: TGF-beta: the missing link in CD4+CD25+ regulatory T cell-mediated immunosuppression. Cytokine Growth Factor Rev 14: 85-89, 2003
- 57) Marleau AM, Sarvetnick N: T cell homeostasis in tolerance and immunity. J Leukoc Biol 78: 575-584, 2005
- 58) Louten J, Boniface K, de Waal Malefyt R: Development and function of TH17 cells in health and disease. J Allergy Clin Immunol 123: 1004-1011, 2009. doi: 10.1016/j.jaci.
- 59) Ricci M, Rossi O, Romagnani S, Del Prete GF: Etiologic factors and pathogenetic aspects of organ-specific autoimmune diseases. Essential role of autoreactive T cells and lymphokine network in the activation of effector systems responsible for tissue lesions. Autoimmunity 2: 331-344, 1989
- 60) Hartwell D, Levine J, Fenton M, Francis C, Leslie C, Beller D: Cytokine dysregulation and the initiation of systemic autoimmunity. Immunol Lett. 1994 Dec;43(1-2):15-21.
- 61) Opdenakker G, Van Damme J: Cytokine-regulated proteases in autoimmune diseases. Immunol Today 15: 103-107, 1994
- 62) Correa SG, Maccioni M, Rivero VE, Iribarren P, Sotomayor CE, Riera CM: Cytokines and the immuneneuroendocrine network: What did we learn from infection and autoimmunity? Cytokine Growth Factor Rev 18: 125-134, 2007
- 63) Korn T, Oukka M, Kuchroo V, Bettelli E: Th17 cells: effector T cells with inflammatory properties. Semin Immunol 19: 362-371, 2007
- 64) Nguyen H, Hiscott J, Pitha PM: The growing family of interferon regulatory factors. Cytokine Growth Factor

Rev 8: 293-312, 1997

- 65) Kröger A, Köster M, Schroeder K, Hauser H, Mueller PP: Activities of IRF-1. J Interferon Cytokine Res 22: 5-14, 2002
- 66) Wu P, Hyodoh F, Hatayama T, Sakaguchi H, Hatada S, Miura Y, Takata-Tomokuni A, Katsuyama H, Otsuki T: Induction of CD69 antigen expression in peripheral blood mononuclear cells on exposure to silica, but not by asbestos/chrysotile-A. Immunol Lett 98: 145-152, 2005
- Wu P, Miura Y, Hyodoh F, *et al.*: Reduced function of CD4+25+ regulatory T cell fraction in silicosis patients. Int J Immunopathol Pharmacol 19: 357-368, 2006
- 68) Murakami S, Nishimura Y, Maeda M, Kumagai N, Hayashi H, Chen Y, Kusaka M, Kishimoto T, Otsuki T: Cytokine alteration and speculated immunological pathophysiology in silicosis and asbestos-related diseases. Environ Health Prev Med 14: 216-222, 2009. doi: 10.1007/s12199-008-0063-8.
- 69) Hayashi H, Maeda M, Murakami S, et al.: Soluble interleukin-2 receptor as an indicator of immunological disturbance found in silicosis patients. Int J Immunopathol Pharmacol; 22:53-62, 2009
- 70) Hayashi H, Miura Y, Maeda M, Murakami S, Kumagai N, Nishimura Y, Kusaka M, Urakami K, Fujimoto W, Otsuki T: Reductive alteration of the regulatory function of the CD4(+)CD25(+) T cell fraction in silicosis patients. Int J Immunopathol Pharmacol 23: 1099-1109, 2010
- 71) Aikoh T, Tomokuni A, Matsukii T, Hyodoh F, Ueki H, Otsuki T, Ueki A: Activation-induced cell death in human peripheral blood lymphocytes after stimulation with silicate in vitro. Int J Oncol 12: 1355-1359, 1998
- 72) Otsuki T, Miura Y, Nishimura Y, Hyodoh F, Takata A, Kusaka M, Katsuyama H, Tomita M, Ueki A, Kishimoto T: Alterations of Fas and Fas-related molecules in patients with silicosis. Exp Biol Med (Maywood) 231: 522-533, 2006
- 73) Otsuki T, Sakaguchi H, Tomokuni A, Aikoh T, Matsuki T, Kawakami Y, Kusaka M, Ueki H, Kita S, Ueki A: Soluble Fas mRNA is dominantly expressed in cases with silicosis. Immunology 94: 258-262, 1998
- 74) Otsuki T, Sakaguchi H, Tomokuni A, et al.: Detection of alternatively spliced variant messages of Fas gene and mutational screening of Fas and Fas ligand coding regions in peripheral blood mononuclear cells derived from silicosis patients. Immunol Lett 72: 137-143, 2000
- 75) Tomokuni A, Aikoh T, Matsuki T, Isozaki Y, Otsuki

T, Kita S, Ueki H, Kusaka M, Kishimoto T, Ueki A: Elevated soluble Fas/APO-1 (CD95) levels in silicosis patients without clinical symptoms of autoimmune diseases or malignant tumours. Clin Exp Immunol 110: 303-309, 1997

- 76) Otsuki T, Maeda M, Murakami S, et al.: Immunological effects of silica and asbestos. Cell Mol Immunol 4: 261-268, 2007
- 77) Maeda M, Nishimura Y, Kumagai N, Hayashi H, Hatayama T, Katoh M, Miyahara N, Yamamoto S, Hirastuka J, Otsuki T: Dysregulation of the immune system caused by silica and asbestos. J Immunotoxicol 7: 268-278, 2010. doi: 10.3109/1547691X.2010.512579.
- 78) Lee S, Hayashi H, Maeda M, Chen Y, Matsuzaki H, Takei-Kumagai N, Nishimura Y, Fujimoto W, Otsuki T: Environmental factors producing autoimmune dysregulation--chronic activation of T cells caused by silica exposure. Immunobiology 217: 743-748, 2012. doi: 10.1016/j.imbio.2011.12.009.
- 79) Lee S, Matsuzaki H, Kumagai-Takei N, et al.: Silica exposure and altered regulation of autoimmunity. Environ Health Prev Med 19: 322-329, 2014. doi: 10.1007/s12199-014-0403-9.
- 80) Lo Re S, Dumoutier L, Couillin I, et al.: IL-17Aproducing gammadelta T and Th17 lymphocytes mediate lung inflammation but not fibrosis in experimental silicosis. J Immunol 184: 6367-6377, 2010. doi: 10.4049/ jimmunol.0900459.
- 81) Song L, Weng D, Liu F, Chen Y, Li C, Dong L, Tang W, Chen J: Tregs promote the differentiation of Th17 cells in silica-induced lung fibrosis in mice. PLoS One 7: e37286, 2012. doi: 10.1371/journal.pone.0037286.
- 82) Chen Y, Li C, Weng D, et al.: Neutralization of interleukin-17A delays progression of silica-induced lung inflammation and fibrosis in C57BL/6 mice. Toxicol Appl Pharmacol 275: 62-72, 2014. doi: 10.1016/ j.taap.2013.11.012.
- 83) Song L, Weng D, Dai W, Tang W, Chen S, Li C, Chen Y, Liu F, Chen J: Th17 can regulate silica-induced lung inflammation through an IL-1 β-dependent mechanism J Cell Mol Med 18: 1773-1784, 2014. doi: 10.1111/jcmm.12341.
- 84) Thakur C, Wolfarth M, Sun J, Zhang Y, Lu Y, Battelli L, Porter DW, Chen F: Oncoprotein mdig contributes to silica-induced pulmonary fibrosis by altering balance between Th17 and Treg T cells. Oncotarget 6: 3722-

3736, 2015

- 85) Liu T, Dai W, Li C, Liu F, Chen Y, Weng D, Chen J: Baicalin alleviates silica-induced lung inflammation and fibrosis by inhibiting the Th17 response in C57BL/6 mice. J Nat Prod 78: 3049-3057, 2015. doi: 10.1021/acs. jnatprod.5b00868.
- 86) Wilbourn JD, McGregor DB, Partensky C, Rice JM: IARC reevaluates silica and related substances. Environ Health Perspect 105: 756-759, 1997
- 87) Guha N, Straif K, Benbrahim-Tallaa L: The IARC Monographs on the carcinogenicity of crystalline silica. Med Lav 102: 310-320, 2011
- 88) Jessica M. Mayeux, Rahul D. Pawar, K. Michael Pollard. Silicates and autoimmunity. Otsuki T, Yoshioka Y, Holian A (eds). Biological Effects of Fibrous and Particulate Substances. Part of the series Current Topics in Environmental Health and Preventive Medicine. Tokyo, Japan, Springer Japan. 2015, pp 163-180
- 89) Pernis B, Vigliani EC, Selikoff IJ: Rheumatoid factor in serum of individuals exposed to asbestos. Ann NY Acad Sci 132: 112-120, 1965
- 90) Nigam SK, Suthar AM, Patel MM, Karnik AB, Dave SK, Kashyap SK, Venkaiah K: Humoral immunological profile of workers exposed to asbestos in asbestos mines. Indian J Med Res 98: 274-277, 1993
- 91) Pfau JC, Sentissi JJ, Weller G, Putnam EA: Assessment of autoimmune responses associated with asbestos exposure in Libby, Montana, USA. Environ Health Perspect 113: 25-30, 2005
- 92) Stansfield D, Edge JR: Circulating rheumatoid factor and antinuclear antibodies in shipyard asbestos workers with pleural plaques. Br J Dis Chest 68: 166-170, 1974
- 93) Turner-Warwick M, Parkes WR: Circulating rheumatoid and antinuclear factors in asbestos workers. Br Med J 3: 492-495, 1970
- 94) Zerva LV, Constantopoulos SH, Moutsopoulos HM:

Humoral immunity alterations after environmental asbestos exposure. Respiration 55: 237-241, 1989

- 95) Huuskonen MS, Räsänen JA, Härkönen H, Asp S: Asbestos exposure as a cause of immunological stimulation. Scand J Respir Dis 59: 326-332, 1978
- 96) Pfau JC, Serve K, Woods L, Noonan C. Asbestos exposure and autoimmunity. Otsuki T, Yoshioka Y, Holian A (eds). Biological Effects of Fibrous and Particulate Substances. Part of the series Current Topics in Environmental Health and Preventive Medicine. Tokyo, Japan, Springer Japan. 2015, pp 181-194
- 97) Ferro A, Zebedeo CN, Davis C, Ng KW, Pfau JC. Amphibole, but not chrysotile, asbestos induces anti-nuclear autoantibodies and IL-17 in C57BL/6 mice. J Immunotoxicol 11: 283-290, 2014. doi: 10.3109/1547691X.2013.847510.
- 98) Serve KM, Black B, Szeinuk J, Pfau JC: Asbestosassociated mesothelial cell autoantibodies promote collagen deposition in vitro. Inhal Toxicol 25: 774-784, 2013. doi: 10.3109/08958378.2013.848249.
- 99) Zebedeo CN, Davis C, Peña C, Ng KW, Pfau JC: Erionite induces production of autoantibodies and IL-17 in C57BL/6 mice. Toxicol Appl Pharmacol 275: 257-264, 2014. doi: 10.1016/j.taap.2014.01.018.
- 100) Pfau JC, Serve KM, Noonan CW: Autoimmunity and asbestos exposure. Autoimmune Dis 2014:782045, 2014. doi: 10.1155/2014/782045.
- 101) Mihai C, Tervaert JW: Anti-endothelial cell antibodies in systemic sclerosis. Ann Rheum Dis 69: 319-324, 2010. doi: 10.1136/ard.2008.102400.
- 102) Savage CO, Williams JM: Anti-endothelial cell antibodies in vasculitis. J Am Soc Nephrol 18: 2424-2426, 2007
- 103) Bordron A, Revelen R, Youinou P: Anti-endothelial cell autoantibodies and systemic disease. Isr Med Assoc J 2: 544-549, 2000